

References

Amer, B.M.A., Hossain, M.A., Gottschalk, K. (2010). Design and performance evaluation of a new hybrid solar dryer for banana, *Energy Conversion and Management* 51; 813–820.

An introduction about coconut industry 1997, Copra and coconut oil industry, by Marketing development and research division, Coconut development authority, Sri Lanka.

Arnold, R. Elepano, Karen, T. Satairapan, (2001), A solar-biomass dryer for pineapple, A paper presented at the 51st Philippine Society of Agricultural Engineers National Convention held in Cebu Plaza Hotel, Cebu City, 23 to 27 April 2001.

Cigdem Tiris, Necdet Ozbalta, Mustafa Tiris, Ibrahim Dincer, (1995), Thermal performance of a new solar air heater, *International Communications in Heat and Mass Transfer*, Vol. 22, No. 3, pp. 411-423,

Coconut oil production 1997, Copra and coconut oil industry, by Marketing development and research division, Coconut development authority, Sri Lanka.

Copra industry in Sri Lanka 1997, Copra and coconut oil industry, by Marketing development and research division, Coconut development authority, Sri Lanka.

Dilip Jain , (2005), Modeling the system performance of multi-tray crop drying using an inclined multi-pass solar air heater with in-built thermal storage, *Journal of Food Engineering* 71. 44–54.

Ebru Kavak Akpinar, Fatih Koçyigit,(2010), Experimental investigation of thermal performance of solar air heater having different obstacles on absorber plates, *International Communications in Heat and Mass Transfer* 37. 416–421.

Fadhela, S. Koolia, Farhata, A., Bellghithb, A., (2005), Study of the solar drying of grapes by three different Processes, *Desalination* 185 (2005) 535–541.

Fereidoon Shahidi. (2005), *Bailey's Industrial Oil and Fat Products*, Sixth Edition, Six Volume Set. 123-147.

Gauhar, A. Mastekbayeva, Chandika, P. Bhatta, Augustus, M., Leon and Kumar, S., (1999), Experimental studies on a hybrid dryer, Paper presented at the ISES 99 Solar World Congress, Israel, 4-9 July 1999, Energy Program, Asian Institute of Technology, P.O. Box 4, Klong Luang, Pathumthani 12120, Thailand

Gopala Krishna, A.G., Gaurav Raj, Ajit Singh Bhatnagar, Prasanth Kumar, P.K. and Preeti Chandrashekar, (2010), *Coconut Oil: Chemistry, Production and Its Applications - A Review*. *Indian Coconut Journal*, 15-27.

- Hachemi, A., (1997), Thermal heat performance enhancement by interaction between the radiation and convection in solar air heaters, *Renewable Energy*, Vol. 12, No. 4, pp. 419-433, 1997.
- Khalil, E.J. Al-Juamili, Abdul Jabbar, N. Khalifa, Tadahmun, A. Yassen, (2007), Testing of the performance of a fruit and vegetable solar drying system in Iraq, *Desalination* 209 163–170.
- Karim, M.A., Hawlader, M.N.A., (2003), Development of solar air collectors for drying applications, *Energy Conversion and Management* 45. 329–344.
- Krishna Raghavan,(2010), Biofuels from coconuts, viewed 23 February 2011, http://www.fact-foundation.com/en/Knowledge_and_Expertise/Media_Library/Full_Library?session=lu0svm0sgap8r92q9h2nfcv9v6.html
- Lyes Bennamoun, Azeddine Belhamri, (2002), Design and simulation of a solar dryer for agriculture products, *Journal of Food Engineering* 59. 259–266,
- Mcdoom, I.A., Ramsaroop, R., Saunders, R., Tang Kai, A., (1999), Optimization of Solar Crop Drying, *Renewable Energy* 16 (1999) 749-752
- Madhlopa, G. Ngwalo (2007), Solar dryer with thermal storage and biomass-backup heater, *Solar Energy* 81. 449–462,
- Mohanraj, M., Chandrasekar, P. (2008), Comparison of drying characteristics and quality of copra obtained in a forced convection solar dryer and sun drying. *Journal of scientific and industrial research* 67. 381-385.
- Mohanraj, M., Chandrasekar, P. (2008), Drying of copra in a forced convection solar dryer, *Biosystems Engineering*, 99, 604 – 607
- Mumba, J., (1995), Design and Development of a Solar Grain Dryer Incorporating Photovoltaic Powered Air Circulation, *Energy Conuers. Mgmt Vol. 37, No. 5*, pp. 615-621,
- Mustafa Aktaş, İlhan Ceylanb, Sezayi Yilmazb, (2009), Determination of drying characteristics of apples in a heat pump and solar dryer, *Desalination* 239. 266–275.
- Peiris, T. S. G., Sri Lankans are using more palm oils than coconut oil, Viewed 19 March 2011, <http://www.cri.lk/articles.html>.
- Roberto C. Guarte, Werner Mfihlbauer, Manfred Kellert, (1996), Drying characteristics of copra and quality of copra and coconut oil, *Postharvest Biology and Technology* 9, 361-372,
- Rodrigo, M. C. P., Arnarasiriwardene, B. L. and Samarajeewa, U. (1996), Some observations on copra drying in Sri Lanka, *COCOS* (1996) 11. 21 -31.

Sarsavadia, P.N., (2007), Development of a solar-assisted dryer and evaluation of energy requirement for the drying of onion, *Renewable Energy* 32. 2529–2547.

Satter, M. A. (2001), Optimization of Copra Drying Factors by Taguchi Method, 4th International Conference on Mechanical Engineering, December 26-28. Dhaka, Bangladesh/pp. III 23-27.

Satter, M. A. (2003), Design and development of a portable copra dryer, Proceedings of the International Conference on Mechanical Engineering. 26- 28 December 2003, Dhaka, Bangladesh.

Shanmugama, V., Natarajanb, E., (2006), Experimental investigation of forced convection and desiccant integrated solar dryer, *Renewable Energy* 31 1239–1251.

Thanaraj, T., Dharmasena, N.D.A., Samarajeewa, U. (2004), Development of a rotary solar hybrid dryer for small scale copra processing, *Tropical Agricultural Research*, 16. 305-315.

Thiruchelvam Thanaraj, Nimal, D.A. Dharmasena, Upali Samarajeewa (2007), Comparison of quality and yield of copra processed in CRI improved kiln drying and sun drying, *Journal of Food Engineering* 78 1446–1451,

Thiruchelvam Thanaraj, Nimal, D.A. Dharmasena, Upali Samarajeewa (2007), Comparison of drying behavior, quality and yield of copra processed in either a solar hybrid dryer on in an improved copra kiln. *International journal of food science and technology* 42. 125-132.

Turhan Koyuncu, (2006), Performance of various designs of solar air heaters for crop drying applications, *Renewable Energy* 31 (2006) 1073–1088.

Wijerathna, M. C. P., Samarajeewa, U., Rodrigo, M.C.P. (1996), *J. Natn. Coun. Sri Lanka* 24. 285-297.

Bibliography

Atul Sharma, Chen, C.R., Nguyen Vu Lan, (2009), Solar-energy drying systems: A review, *Renewable and Sustainable Energy Reviews* 13. 1185–1210.

Ayyappan, S. and Mayilsami, K. (2010), Experimental investigation on a solar tunnel dryer for copra drying. *Journa of scientific and industrial research* 69. 635-638.

Codex Stan 210-1999, (1999), Page 11, viewed 21 August 2011, http://www.codexalimentarius.net/web/standard_list.do?lang=en for RI

Ekechukwua, O.V., Nortonb, B., (1999), Review of solar-energy drying systems II: an overview of solar drying technology, *Energy Conversion & Management* 40 615±655.

Fudholi, Sopian, K., Ruslan, M.H., Alghoul, M.A., Sulaiman, M.Y., (2010), Review of solar dryers for agricultural and marine products, *Renewable and Sustainable Energy Reviews* 14, 1–30

Kevin, L. Goodner, and Daniel, J. Wampler, (2009), Sensus Technical Note (SEN-TN-0026), 7255 Hamilton Enterprise Park Drive, Hamilton, OH 45011.

Michael, W. Bassey, (1985), Design and performance of a hybrid crop dryer using solar energy and sawdust, *Proceedings of international conference*. 22-28 June.

Smarajeewa, U., Wei, C. I., Fernando, S. Y., Ahmed, E.M. (1991), Conditions necessary for inactivation of Aflatoxin B1 and loss of mutagenicity in copra meal on chlorine gas treatment. *J. Natn. Sci. Coun. Sri Lanka* 25, 19-32.

Tesha,(2006), Utilization of brine water for copra drying in lahendong geothermal field, *Proceedings at geothermal training programme, Orkustofnun, Grensásvegur 9, IS-108 Reykjavík, Iceland. Report 20, 453-470.*

Appendix A

Table A1: Manufacture of copra, coconut oil in Sri Lanka (CDA statistics)

Year	Copra (MT)	Coconut Oil (MT)
1975	203082	120263
1976	162818	100196
1977	79885	49160
1978	130145	80089
1979	163145	100397
1980	101619	62535
1981	125083	75668
1982	170397	102485
1983	138408	82884
1984	62295	36824
1985	218456	129807
1986	242616	143300
1987	125372	71622
1988	61987	34548
1989	129528	74633
1990	127806	74804
1991	60568	32556
1992	54956	30219
1993	40685	22000
1994	103985	60000
1995	112643	64515
1996	74468	41000
1997	67348	36185
1998	70272	38000
1999	64602	36000
2000	82062	44407
2001	112996	64320
2002	59033	30100
2003	49466	21000
2004	37081	25000
2005	37419	20000

Table A2: Edible oil consumption pattern in Sri Lanka (Peiris, T. S. G., n.d.)

Year	CNO (MT)	Non-CNO (MT)	Rate of oil consumption (Bottle/person/year)		
			CNO	Non-CNO	Both
1989	44965	21000	4.11	1.92	6.03
1990	62580	19712	5.66	1.78	7.44
1991	31507	27231	2.81	2.43	5.23
1992	30234	40012	2.67	3.53	6.20
1993	48800	42235	4.253	3.68	7.94
1994	56121	55276	4.83	4.75	9.58
1995	55562	36965	4.71	3.14	7.85
1996	38688	73461	3.25	6.16	9.41
1997	50915	85023	4.22	7.04	11.26
1998	35498	90498	2.91	7.42	10.32
1999	31503	117284	2.55	9.48	12.02
2000	39751	90079	3.16	7.16	10.32
2001	61106	72662	5.02	5.97	10.99
2002	52896	142480	4.28	11.53	15.81
2003	82613	119502	6.60	9.55	16.15
2004	1 26105	111699	2.06	8.82	10.88

Appendix B

Table B1: Characteristics of coconut oil

Characteristic	Edible type						Non edible type	Method of Test
	Refined, bleached and deodorized coconut oil	Refined and bleached coconut oil	White coconut oil	Coconut oil	Paring coconut oil	Virgin coconut oil	Industrial coconut oil	
Colour 25-mm cell on the Lovibond colour scale expressed in Y+5R, not deeper than	2	2	4	5	5	1	11	SLS 313
Relative density at 30°C/30°C	0.915 to 0.920	0.915 to 0.920	0.915 to 0.920	0.915 to 0.920	0.915 to 0.920	0.915 to 0.920	0.915 to 0.920	
Relative density at 40°C	1.448 to 1.4492	1.448 to 1.4492	1.448 to 1.4492	1.448 to 1.4492	1.448 to 1.4492	1.448 to 1.4492	1.448 to 1.4492	
Moisture & other matter Volatile at 105°C, max.	0.1	0.1	0.4	0.4	0.4	0.5	0.5	
Insoluble impurities, per Cent by mass, max	0.05	0.05	0.05	0.05	0.05	0.05	0.05	
Free fatty acids, as lauric Acid, per cent by mass	0.1(max.)	0.1(max.)	0.8(max.)	0.8(max.)	0.8(max.)	0.2(max.)	1.0 to 5.0(or as traded)	
Iodine value	7.5 to 9.5	7.5 to 9.5	7.5 to 11.0	7.5 to 11.0	9.0 to 18.0	6.0 to 11.0	7.5 to 18.0	
Saponification value	255(min.)	255(min.)	248 to 264	248 to 264	248 to 264	255(min.)	248 to 264	
Unsaponifiable matter, per cent by mass, max	0.5	0.8	0.8	0.8	0.8	0.2	0.8	
Mineral acidity	Nil	Nil	Nil	Nil	Nil	Nil	Nil	

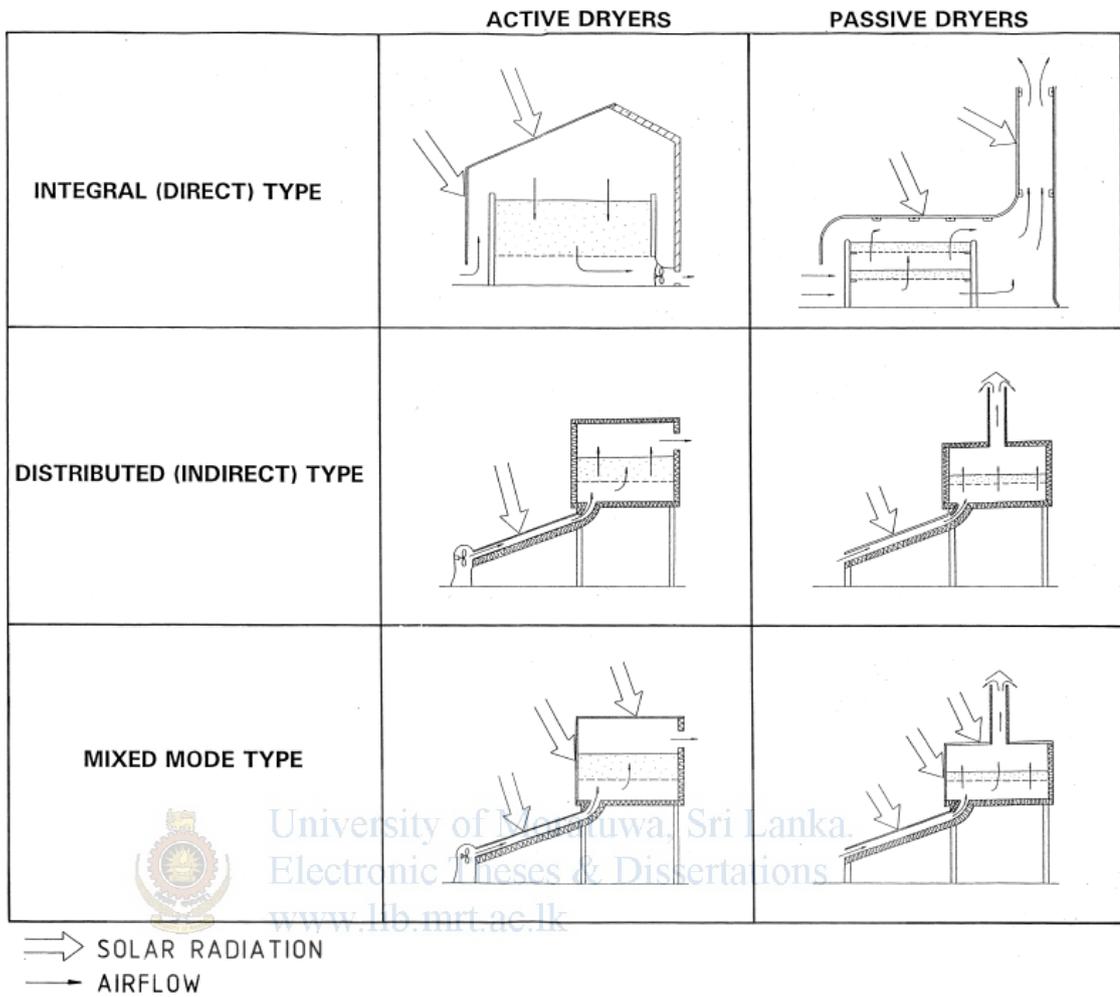


Figure B1 Typical solar energy drier designs
 (Ekechukwua, O.V. and Nortonb, B., 1999)

Appendix C

1. Details of equipments

Equipment	Model Name	Technical Details
Luxmeter	Testo 545	Range 0-100000 lux Resolution :1 lux (0-32000 lux) Resolution : 10 lux (0-100000 lux)
Velometer	Alnor	Range of pressure = 0 to 10 inches H ₂ O Resolution: = 0.01 inches H ₂ O Range of flow rate: = 0-10000 ft/min Resolution: = 20 ft/min
Oven	Griffin 1/200	Range of temperature = 0°C to 200 °C



(Luxmeter)



(Velometer)



(Oven)

2. Soxhelt Extraction of Coconut Oil

Procedure:

1. Apply a thin layer of grease at all the ground glass joints.
2. Weigh 10 grams of scraped coconut accurately in to the extraction thimble.
3. Introduce 300 ml of solvent to the distillation flask.
4. Fit the condenser on top of the extractor tube, and circulate water.
5. Start heating the flask. Reflux the solvent for about an hour (4 to 5 times).
6. Switch off the heating and allow it to cool. Then remove the thimble from the extractor tube and fit the apparatus back. Distill the solvent to the extractor tube and collect it for reuse. Then weigh the oil left behind in the flask.

Results:

Represent the oil content by weigh percentage on the basis of the scraped coconut used in the experiment.



$$\text{Oil Content} = \frac{\text{Amount of oil extracted}}{\text{Weight of coconut}} \times 100$$

University of Moratuwa, Sri Lanka
Electronic Theses & Dissertations
www.lib.mrt.ac.lk

3. Tests of coconut oil

1. Free Fatty Acid (FFA) content

This test measure the free fatty acid content by a titration with ethonolic KOH and gives the value as a percentage. FFA % is defined as the number of KOH required to neutralize the free fatty acid in 1g of oil. The content of free fatty acid is increased due to the moisture and sunlight and therefore can be reduced if refrigerated.

Chemicals :

- NaOH or KOH solution – for RBD oil, refined oil and white oil normality of the solution should be 0.1. For industrial oil solution it should be 0.5 N.
- Phenolphthalein indicator

- Ethanol, 95% (V/V), neutral to the phenolphthalein indicator

Testing procedure :

- Correctly weigh out 40g of RBD coconut oil or refined oil (if it is white coconut oil the mass is 20g and if it is industrial oil mass is 10g) to a 250 ml conical flask.
- Add 50ml of the solvent (solvent consists of equal amounts of ethanol and petroleum ether)
- Heat it on a water bath
- Titrate the mixer with 0.1 mol dm^{-3} KOH solution using phenolphthalein as the indicator
- Take the end point of the titration as the colorless oil solution turns to pink color and take the reading.
- The FFA percentage can be measured using the formula given below.

$$\text{FFA}\% = (V * N * 20.0 / (W))$$

Where,

V= Volume of the KOH obtained from the titration

N = Normality of the KOH (OH^- ion concentration – here N = 0.1)

W = Weight of the oil sample (25 g)

2. Iodine value

Iodine value is defined as the number of grams of Iodine which react with 100g of coconut oil. The Iodine value measures the quantity of unsaturated fatty acid by a titration method. The unsaturation is due to the presence of double bonds. Higher the iodine value indicates that there are more unsaturated fatty acids present in the oil.

Testing Procedure (Wij's Method):

- 2 – 2.5 grams of oil is mixed with 15 ml CCL_4 to dissolve the fatty acid on CCL_4 in an iodine flask.

- Then the solution is treated with an 20 ml of Wij's iodine solution and seal the bottle. Wij's iodine solution is a solution of iodine monochloride (ICl) in glacial acetic acid. (Pale yellow color solution)
- Allow the bottle to stand for 30 minutes in dark.
- Next 15 ml of potassium iodide solution and 100 ml of distilled water is added. Unreacted iodine monochloride is reacted with potassium iodide which converts it to iodine.
- This is immediately titrated against 0.1 M sodium thiosulfate solution using phenolphthalein as the indicator. The color change from pale yellow to colorless.
- Add 3 ml of starch. Then the solution turns in to dark blue color.
- The iodine concentration is then determined by titration with sodium thiosulfate
- At the end point the color changes from blue to colourless.
- 1 ml of 0.1 M sodium thiosulfate solution = 0.01269 g of iodine.
- The difference between a control titration and the titration with the fat present multiplied by this factor gives the mass of iodine absorbed by the oil.
- The iodine value can be calculated by the given equation

$$\text{Iodine Value} = \frac{(\text{mL of blank} - \text{mL of sample}) \times N(\text{sodium thiosulfate}) \times 12.69}{\text{Weight of sample (g)}}$$

3. Saponification value

This is the number of KOH needed to neutralize the fatty acids formed by the complete hydrolysis of 1gram of the lipid. This method is used to determine the total acid content, both free and combined, of coconut oil. The combined acids are

primarily esters formed by reaction with the neutral components present in the original oil.

Chemicals:

- KOH 0.5 N solution in 95% ethanol.
- 0.5 N HCl solution
- Phenolphthalein indicator

Procedure:

- Weigh 2 g of oil sample, to the nearest 0.01 g, into a 500ml clean, dry, flat bottomed flask.
- Add 25 ml of 0.5 N ethanolic potassium hydroxide by using a pipette.
- Boil continuously under a Reflux condenser for 60 minutes.
- Titrate between 60 and 70°C with 0.5 N HCl acid using phenolphthalein indicator
- Run a blank in the same manner.

Calculation :

$$\text{Saponification Value} = \{ (A - B) \times N \times 56.1 \} / W$$

Where,

A = HCl volume, for blank in mL

B = HCl volume, for sample in mL

W = weight of sample (dry basis), g

N = normality of HCl solution

56.1 = weight of potassium hydroxide

4. Relative Density

Relative density is the density of the oil with respect to the water at the temperature of 30°C. It is measured by using a 50 ml density bottle.

Procedure :

- Weigh out the dry empty density bottle with stopper correctly to four decimal places. (W1 g)
- Cool or heat the oil until the temperature of the oil reaches 30 °C.
- Then fill the bottle with coconut oil and measure the weight (W2 g). the filling must be done slowly and carefully while holding the bottle in a slanting position where the air bubbles are not taken in with the oil. the froth produced by the air bubbles are carefully removed and the stopper is carefully inserted before measuring the weight.
- The relative density is given by the following equation

$$\text{Relative density} = (W2 - W1) / 50$$

Since 50 ml density bottle is taken the weight of the water that can be filled to the density bottle is equalent to 50 g.

5. Color

Color of coconut oil is measured by using a Lovibond Tintometer in 25 mm cell. Lovibond Tintometer is an automatic instrument and is very easy to use,. There is no need to build up calibration curves as they are already established in the instrument. The menu system guides operators through the selection of operating parameters. Thereafter, measurements are initiated by just a single key press and take less than 25 seconds to complete.

6. Refractive Index

Refractive index of coconut oil is the ratio between the velocities of the life of a definite wave length in vacuum to the velocity in the oil medium. Refractive index is measured using a refractometer.

Procedure :

- Calibrate the refractometer and its accuracy using liquids with knowing refractive index.

- Filter the oil sample through filter paper to remove any impurities and the last traces of moisture. The sample must be completely dry.
- Make certain that the temperature of the refractometer is 40.0°C
- Fill the space between the prisms of the refractometer completely with oil. Close the prisms and tighten firmly with a screw head.
- Allow to stand for 3 minutes so that the sample comes to the temperature of the instrument.
- Adjust the instrument and light to obtain the most distinct reading possible and then determine the refractive index. Take the readings in a temperature of nearly 40.0°C.
- Take several readings and calculate the average of all. After each determination, the prisms of the instrument should be cleaned with a cotton wool soaked with a mixture of ether and dry alcohol.

The refractive index at any other temperature can be calculated from the equation given below.

$$R' = R + 0.0036 * T \text{ (For temperatures higher than } 40.0^{\circ}\text{C)}$$

$$R' = R - 0.0036 * T \text{ (For temperatures lower than } 40.0^{\circ}\text{C)}$$

Here R' = refractive index at a different temperature

R = refractive index at 40.0°C

T = temperature difference in °C

7. Moisture

The moisture content of the oil can be measured by either oven method or using a moisture balance. In the oven method a sample of oil is placed in an oven and heated to 101 – 105 °C and the loss of mass due to the vaporization is calculated.

In the moisture balance method the sample is placed on a moisture balance for 5 – 10 minutes. Then the balance in the instrument is balanced by adjusting the weights thereby the moisture content can be directly obtained.

8. Mineral acidity

Mineral acidity measures the contents of mineral such as HCL and H₂SO₄ acids mixed with oil.

Chemicals:

- NaOH or KOH solution
- Light petroleum (Boiling point 40°C – 60°C)
- Methyl orange indicator

Test Procedure:

- 50g of oil is accurately weighed to a separating funnel
- The oil is washed 3 times from portions of 50 – 60 ml of hot distilled water.
- The hot water washings were combined to another separating funnel.
- The water was allowed to be cooled to the room temperature.
- The traces of fatty acids were removed by allowing it to be extracted to 50ml of light petroleum.
- The wash water was separated out and titrate with 0.01N NaOH or KOH solution using methyl orange as the indicator.

$$\text{Mineral Acidity} = 100 * V / m$$

Here V = titration volume of NaOH or KOH in ml, m = mass of the oil taken in grams

Here the mineral acidity is obtained as milliliters of 0.01 N acids per 100 grams of oil.

9. Unsaponifiable Matter

This measures the other contents in coconut oil such as proteins, carbohydrates, alcohol, sterol etc.

Chemicals:

- 95% Ethanol
- Ethonolic KOH solution
- Diethyl Ether (relative density 0.72 – 0.724)
- HCl acid (relative density .1.6 – 1.18)

- 0.5N KOH solution
- Phenolphthalein indicator
- Acetone
- 0.1N Ethanolic NaOH solution

Procedure:

- 2 – 2.2 grams of oil is accurately weighed and transferred to a 250ml flask.
- 25ml of Ethanolic NaOH solution was added and to the oil and heated and refluxed with frequent swirling on a boiling water bath for 1 hour
- The ethanolic soap solution was transferred to a separating funnel and rinsed with 50ml of water for several times.
- Allowed to be cooled to the room temperature.
- Flask is rinsed with 50ml of ether and transferred to the separating funnel.
- Separating funnel is stoppered and shake vigorously.
- The solution was allowed to be separated and clarified.
- The lower aqueous ethanolic solution was transferred to a flask.
- The ethereal extract was poured from the top of the separating funnel into a second 250ml separating funnel which contains 20ml of water..
- The aqueous ethanolic solution was transferred to the first separating funnel from the flask. The extraction was repeated twice using 50ml of ether while adding the extracted ether layer to the second separating funnel..
- If a filter has been used, the filter was also washed with ether and added to the second separating funnel
- The liquid in the second separating funnel was swirled without shaking and allowed to be separated and the lower aqueous layer was discarded.
- The ethereal layer was washed out twice using further 20ml of water portions and discarding the lower aqueous layer.
- 20ml of 0.5N KOH solution was added to the ethereal solution and was shake vigorously. It was allowed to be separated and the aqueous layer was removed. Same procedure was followed using 20ml water.

- The washing with alkali and then with water was followed more for two times.
- The washing with water was continued until the residue water become neutral to the phenolphthalein indicator.
- Ethereal solution was transferred to a weighed flask and most of the ether was distilled off.
- 2 – 3 ml of acetone was added and the removal of the solvent was continued
- The flask was then dried out until a constant mass was obtained. The temperature must not exceed 80°C.
- The residue was dissolved in 10ml of freshly boiled and neutralized ethanol and it was titrated with 0.1N ethonolic NaOH using phenolphthalein as indicator.
- If the volume of 0.1N ethonolic NaOH required is less than 0.1ml, the mass of the residue can be taken as the mass of Unsaponifiable matter.
- If the volume exceeds 0.1ml the test have to be rejected and repeated.

The Unsaponifiable matter % by mass = $(M_1/M_0) * 100$

Here, M_1 = mass of residue in grams

M_0 = mass of oil taken in grams

10. Insoluble impurities

Insoluble impurities are defined as the dirt and other foreign matter insoluble in light petroleum. It is expressed as a percentage by mass.

Procedure :

- A weighed quantity of 20ml – 50ml of oil at a temperature below 60°C is filtered through an ashless open texture filter paper which was previously dried in an oven at 105 °C and weighed in a stoppered weighing bottle.
- If the oil is slow in filtering the oil has to be dialuted in light petroleum (the boiling point of the petroleum should be between 40 °C – 60 °C) before filtration

- In a continuous extraction apparatus, the filter paper containing impurities is extracted using light petroleum.
- After a complete extraction, the filter paper and the contents are dried in an oven at 98 °C - 100 °C and was reweighed in a stoppered weighing bottle until the mass is constant.

Total impurities % by mass = $(M3 - M2) / M1 * 100$

Here,

M1 = mass of oil taken in grams

M2 = mass of weighing bottle and the dry filter paper in grams

M3 = mass of weighing bottle and the dry filter paper containing impurities in grams

For all the above mentioned tests there is a Sri Lankan Standard (SLS) which express the critical value. The table below indicates those critical values of refined bleached deodorized oil, refined bleached oil, white coconut oil, paring oil, virgin coconut oil and the industrial coconut oil.



University of Moratuwa, Sri Lanka.
Electronic Theses & Dissertations
www.lib.mrt.ac.lk

Appendix D

1. Calculation of thermal efficiency

Colorific value of firewood	= 14700 KJ/kg
Calorific value of coconut shell	= 12900 KJ/kg
Capacity of blower	= 0.746 Kw
Latent heat of vaporization of water	= 2270 kJ/kg

a) Experiment 1 (Multi bed experiment)

Compartment 1

Amount of final dried product at the (m_f)	= 11.94 kg
Initial moisture content of coconut (M_0)	= 49.13 %
Final moisture content of coconut (M_f)	= 6.6 %
Oil content of copra	= $11.94 \times (1 - 0.066)$
	= 11.152 kg
Initial weight of coconut without shells	= $11.152 \times 1 / (1 - 0.4913)$
	= 21.922 kg
Weight loss during drying	= $21.922 - 11.94$
	= 9.982 kg

Compartment 2

Amount of final dried product at the (m_f)	= 12.11 kg
Initial moisture content of coconut (M_0)	= 48.62 %
Final moisture content of coconut (M_f)	= 6.91 %
Oil content of copra	= $12.11 \times (1 - 0.0691)$
	= 11.273 kg
Initial weight of coconut without shells	= $11.273 \times 1 / (1 - 0.462)$
	= 21.941 kg
Weight loss during drying	= $21.941 - 12.11$
	= 9.831 kg

Compartment 3

Amount of final dried product at the (m_f)	= 12.37 kg
Initial moisture content of coconut (M_0)	= 48.82 %
Final moisture content of coconut (M_f)	= 7.11 %
Oil content of copra	= $12.37 \times (1 - 0.071)$ = 11.49 kg
Initial weight of coconut without shells	= $11.49 \times 1 / (1 - 0.4882)$ = 22.451 kg
Weight loss during drying	= $22.451 - 12.37$ = 10.081 kg
Initial Surface moisture in coconut halves	= 7.12 kg
Total weight loss during drying	= 37.006 kg
Amount of energy used to evaporate the moisture	= 37.006×2270 kJ = 84003.62 kJ
Amount of firewood used for drying coconut	= 51 kg
Amount of coconut shells used for drying coconut	= 25 kg
Amount of energy supplied by biomass	= $51 \times 14700 + 25 \times 12900$ = 1072200 kJ
Number of operating hours of the blower	= 45
Energy consumed by the blower	= $45 \times 0.746 \times 3600$ = 120852 kJ
Total energy utilized for drying of coconut	= $1072200 + 120852$ = 1193052 kJ
Thermal efficiency of the drier	= $(84003.62 / 1193052) \times 100$ = 7.07 %

b) Experiment 2 (Single bed experiment)

Amount of final dried product at the (m_f)	= 12.06 kg
Initial moisture content of coconut (M_0)	= 49.81 %

Final moisture content of coconut (M_f)	= 7.00 %
Oil content of copra	= $12.06 \times (1 - 0.070)$ = 11.216 kg
Initial weight of coconut without shells	= $11.216 \times 1 / (1 - 0.4981)$ = 22.347 kg
Weight loss during drying	= $22.347 - 12.06$ = 10.287 kg
Initial Surface moisture in coconut halves	= 2.192 kg
Total weight loss during drying	= 12.479 kg
Amount of energy used to evaporate the moisture	= 12.479×2270 kJ = 28327.33 kJ
Amount of firewood used for drying coconut	= 34 kg
Amount of coconut shells used for drying coconut	= 8 kg
Amount of energy supplied by biomass	= $34 \times 14700 + 8 \times 12900$ = 603000 kJ
Number of operating hours of the blower	= 37
Energy consumed by the blower	= $37 \times 0.746 \times 3600$ = 99367.2 kJ
Total energy utilized for drying of coconut	= $603000 + 99367.2$ = 702367.2 kJ
Thermal efficiency of the drier	= $(28327.33 / 702367.2) \times 100$ = 4.05 %

c) Experiment 3 (Single bed experiment)

Amount of final dried product at the (m_f)	= 12.24 kg
Initial moisture content of coconut (M_0)	= 50.41 %
Final moisture content of coconut (M_f)	= 6.75 %
Oil content of copra	= $12.24 \times (1 - 0.0675)$ = 11.414 kg

Initial weight of coconut without shells	= $11.414 \times 1 / (1-0.5041)$
	= 23.016 kg
Weight loss during drying	= $23.016 - 12.24$
	= 10.776 kg
Initial Surface moisture in coconut halves	= 2.370 kg
Total weight loss during drying	= 13.146 kg
Amount of energy used to evaporate the moisture	= 13.146×2270 kJ
	= 29841.42kJ
Amount of firewood used for drying coconut	= 36 kg
Amount of coconut shells used for drying coconut	= 8 kg
Amount of energy supplied by biomass	= $36 \times 14700 + 8 \times 12900$
	= 632400 kJ
Number of operating hours of the blower	= 35
Energy consumed by the blower	= $35 \times 0.746 \times 3600$
	= 93996 kJ
Total energy utilized for drying of coconut	= $632400 + 93996$
	= 726396 kJ
Thermal efficiency of the drier	= $(29841.42 / 726396) \times 100$
	= 4.18 %

2. Calculation of Specific moisture evaporation rate (SMER)

a) Experiment 1 (Multi bed experiment)

Amount of water evaporated from the drying of coconut	= 37.006 kg
Amount of energy utilized for the drying of coconut	= 1193052 kJ
SMER	= $1193052 \text{ kJ} / 37.006 \text{ kg}$
	= 32.11 GJ/kg

b) Experiment 2 (Single bed experiment)

Amount of water evaporated from the drying of coconut = 12.479 kg
Amount of energy utilized for the drying of coconut = 702367.2kJ
SMER = 702367.2 / 12.479 kg
= 55.99 GJ/kg

c) Experiment 3 (Single bed experiment)

Amount of water evaporated from the drying of coconut = 13.146 kg
Amount of energy utilized for the drying of coconut = 726396 kJ
SMER = 726396 / 13.146 kg
= 54.27 GJ/kg



University of Moratuwa, Sri Lanka.
Electronic Theses & Dissertations
www.lib.mrt.ac.lk